

Preliminary communication

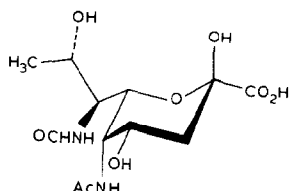
The structure of the *Pseudomonas aeruginosa* immunotype 6 O-antigen: isolation and identification of 5-acetamido-3,5,7,9-tetra-deoxy-7-formamido-L-glycero-L-manno-nonulosonic acid

YURIY A. KNIREL, NINA A. KOCHAROVA, ALEXANDER S. SHASHKOV, and NIKOLAY K. KOCHETKOV

N. D. Zelinsky Institute of Organic Chemistry, Academy of Sciences of the U.S.S.R., Moscow (U.S.S.R.)

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Derivatives of 5,7-diamino-3,5,7,9-tetra-deoxynonulosonic (pseudaminic) acid have been found¹ in some bacterial lipopolysaccharides (LPS), and one of them has been identified as 5-acetamido-3,5,7,9-tetra-deoxy-7-[(*R*)-3-hydroxybutyramido]-L-glycero-L-manno-nonulosonic acid². We now report the identification of 5-*N*-acetyl-7-*N*-formyl-pseudaminic acid (**1**) which is the first representative of these new sialic acid-like sugars isolated in a free state. The structure of the O-specific side-chain of *P. aeruginosa* immunotype 6 LPS containing **1** has also been established.



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The LPS was isolated from dry bacterial cells by phenol–water extraction³. The ¹³C-n.m.r. data indicated it to have the same monosaccharide composition as the LPS from *P. aeruginosa* O5a,b,d, including *N*-acetyl-*N'*-formylpseudaminic acid, xylose, and 2-acetamido-2,6-dideoxygalactose¹. The last two sugars, which were found earlier⁴ to be the components of this LPS, were isolated after mild acid hydrolysis of the LPS (1% HOAc, 100°, 1.5 h) followed by solvolysis with HF (20°, 3 h) and were identified as D isomers on the basis of their optical rotations. Both aldoses are present in the pyranoid form and are β (105.5 and 99.4 p.p.m., both C-1, ¹J_{C,H} 162 and 163 Hz*, respectively)⁵.

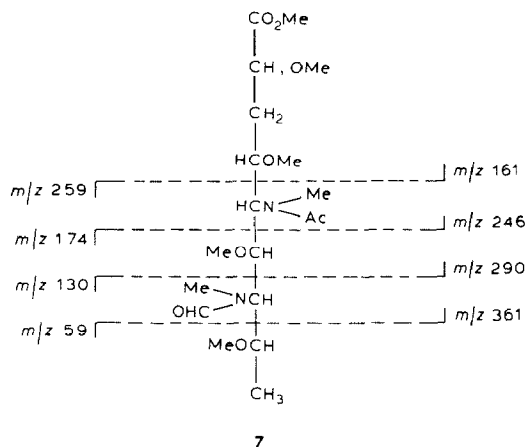
On solvolysis with HF in methanol^{2,6} (20°, 2 h), the LPS gave the methyl glycosides of disaccharides 2 and 3, as well as of trisaccharides 4–6: α-PseN₂AcFm-(2→4)-β-D-Xylp-(1→OMe) (2), α-PseN₂AcFm-(2→4)-α-D-Xylp-(1→OMe) (3), α-PseN₂AcFm-(2→4)-β-D-Xylp-(1→3)-β-D-FucNAcp-(1→OMe) (4), α-PseN₂AcFm-(2→4)-β-D-Xylp-(1→3)-α-D-FucNAcp-(1→OMe) (5), and α-PseN₂AcFm-(2→4)-β-D-Xylp-(1→3)-β-D-FucNAcf-(1→OMe

*Determined from the gated-decoupling spectrum.

(6), where PseN_2AcFm is 1. The oligosaccharides were identified by using ^{13}C -n.m.r. spectroscopy, the spectra being interpreted by comparison with each other and with those of the LPS and 1 (see below).

Analysis of the glycosidation effects in the spectra gave information on the positions of the glycosidic linkages⁷. Thus, in the spectrum of 2, the signals for C-3,5 of the xyloside residue were displaced by -1.6 and -2.0 p.p.m., respectively, as compared with those for methyl β -D-xylopyranoside⁸, which is characteristic of substitution of this sugar in position 4. The small α -effect ($+1.4$ p.p.m.) for the signal of C-4 of xylose also accords with ketosidic substitution at O-4. 2-Acetamido-2,6-dideoxygalactose is glycosidated at O-3, as follows from the low-field position of the signal for C-3 at 81.3 p.p.m. in the spectrum of the LPS. Finally, comparison of the chemical shifts for the signals of C-3,4,5 of pseudaminic acid in 2 (36.8, 66.3, and 49.9 p.p.m.) and the LPS (35.4, 72.4, and 46.5 p.p.m.) showed this sugar to be substituted at position 4 in the LPS and to occupy the non-reducing end in the oligosaccharide. These data also revealed the sequence of the monosaccharides in the repeating unit.

Formation of the oligosaccharides 2–6 proved the glycosidic linkage of pseudaminic acid to be more stable towards HF than those of xylose and 2-acetamido-2,6-dideoxygalactose. In contrast, this linkage was extremely labile towards acid, and mild hydrolysis (1% HOAc, 100° , 2 h) of 2–6 yielded 1 which was isolated by ion-exchange chromatography on DEAE-Trisacryl M followed by preparative p.c. (ethyl acetate–pyridine–acetic acid–water 5:5:1:3, R_{NeuNAc} 1.0). Compound 1 had $[\alpha]_{\text{D}}^{20} -38^\circ$ (water). N.m.r. data: ^1H , δ 1.08 (d, 3 H, $J_{8,9}$ 6.6 Hz, H-9), 1.77 (dd, 1 H, $J_{3a,3e}$ 13.5, $J_{3a,4}$ 12.3 Hz, H-3a), 1.89 (dd, 1 H, $J_{3e,4}$ 5.5 Hz, H-3e); ^{13}C , 177.3, 97.6, 35.8, 66.7, 49.9, 71.1, 52.7, 67.6, and 16.3 p.p.m. for C-1,2,3,4,5,6,7,8,9, respectively. Reduction of 1 with NaBH_4 and methylation⁹ of the product gave 7. Analysis of the mass spectrum of 7 revealed the positions of the amino groups, the deoxy groups, and the *N*-acyl substituents (acetyl at N-5 and formyl at N-7).



The H,H coupling constants $J_{3a,4}$ 12.3, $J_{4,5}$ 4, and $J_{5,6}$ 2 Hz for 1 indicated an axial, equatorial orientation of H-4,5 and thus proved the *lyxo* configuration of the fragment C-4,5,6. The similar large $J_{6,7}$ values (10.6 and 10.0 Hz, respectively) for 1 and the

derivative of pseudaminic acid from *P. aeruginosa* O10a O-specific polysaccharide¹ proved the C-6,7 fragment in each sugar to have the *erythro* configuration² (a $J_{6,7}$ value of 1.2 Hz has been reported¹⁰ for *N*-acetylneuraminic acid in which this fragment has the *threo* configuration). The *erythro* configuration of the C-7,8 fragment followed from the relatively high-field position (16.3 p.p.m.) of the signal for C-9 of 1, whereas C-9 of the *threo* isomer should resonate near 20 p.p.m. (*cf.* the data¹¹ for C-4 of *N*-acetylallothreonine and *N*-acetylthreonine). Thus, 1 has the same general configuration as the *L-glycero-L-manno* isomer in the *P. aeruginosa* O10a O-specific polysaccharide², which is confirmed also by the similar ¹³C chemical shifts for the signals of C-3,4,5,7,8,9 of both sugars (*cf.* the data in *ref.* 1).

The relative absolute configuration of 1 was established by ¹³C-n.m.r. spectroscopy using the known regularities for glycosidation effects¹². The situation in the disaccharide unit β -D-FucNAcp-(1 \rightarrow 4)-PseN₂AcFm is favorable because of the relatively small α -effects for C-1 of 2-acetamido-2,6-dideoxy- β -D-galactose and for C-4 of pseudaminic acid (+3.3 and +6.0 p.p.m., respectively), as well as the relatively large (by module) β -effect (-3.4 p.p.m.) for C-5 of the latter sugar (the effects were determined by comparison of the ¹³C-n.m.r. data for the LPS and the corresponding free sugars). These data are characteristic of a disaccharide unit β -D-Hexp-(1 \rightarrow 3)-L-Gal¹², the lack of a hydroxyl group at C-3 of pseudaminic acid (corresponding to C-2 of galactose) thus being of little significance. Therefore, the fragment C-4,5,6 of 1 is homomorphic with the fragment C-3,4,5 of L-galactose (but not of D-galactose); hence the new sugar has the *L-glycero-L-manno* configuration.

The relatively small difference (\sim 0.6 p.p.m.) between the chemical shifts of the signals for H-3e and H-3a of pseudaminic acid in 2-6 is characteristic of the equatorial orientation of the carboxyl group¹³ (*cf.* the difference of 0.9 p.p.m. for the pseudaminic acid having an axial carboxyl group in the *P. aeruginosa* O10a O-specific polysaccharide¹). Therefore, this sugar is α in 2-6 in accordance with the rules of carbohydrate nomenclature¹⁴. The higher-field position (\sim 72 p.p.m.) of the signal for C-6 of pseudaminic acid in 2-6, as compared with that (75 p.p.m.) in the *P. aeruginosa* O10a polysaccharide¹, confirmed this conclusion¹⁵. There is only one series of signals for one of the anomers in the ¹H- and ¹³C-n.m.r. spectra of 1, and since the chemical shifts for the signals of C-6 in 1 and in 2-6 are similar, cleavage of the glycosidic linkage of pseudaminic acid was not accompanied by anomerisation.

Thus, the repeating unit of the polysaccharide chain of *P. aeruginosa* immunotype 6 LPS is \rightarrow 4)- α -PseN₂AcFm-(2 \rightarrow 4)- β -D-Xylp-(1 \rightarrow 3)- β -D-FucNAcp-(1 \rightarrow .

According to the ¹³C-n.m.r. data, the O-specific polysaccharide of *P. aeruginosa* O5a,d (Lányi classification) also has this structure, whereas those of the serotypes O5a,b,d and O5a,b,c differ by the presence of an *O*-acetyl group at position 4 of the 2-acetamido-2,6-dideoxygalactose residue. Also, the polysaccharide of serotype O5a,b,c has an (*R*)-3-hydroxybutyryl group at N-5 of pseudaminic acid instead of an acetyl group¹.

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